

Taxonomy and systematics

Discovery of the type material of *Polynoe fasciculosa* (Annelida, Polychaeta, Polynoidae)

Descubrimiento del material tipo de Polynoe fasciculosa (Annelida, Polychaeta, Polynoidae)

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Abstract

Émile Blanchard described 3 polynoid species from Chile: *Polynoe chilensis*, *P. fasciculosa*, and *P. virens*. The type material of *P. chilensis* was found in the Muséum National d'Histoire Naturelle, Paris, but the 2 other species were regarded as indeterminable, and without type specimens. During a research visit in Paris, the type specimens of *P. fasciculosa* were found, and together with some non-type specimens deposited in the Los Angeles County Museum of Natural History, the species is regarded as a senior synonym of *Lepidonotus arenosus* Ehlers, 1901, by having 12 pairs of elytra, lateral antennae terminal, elytra fimbriate with macro- and microtubercles globular to polyhedric, and neurochaetae barely bidentate, with accessory denticles often eroded. However, *P. fasciculosa*, as a junior homonym of *P. fasciculosa* Grube, 1840, or newly combined as *L. fasciculosus*, is regarded as a *nomen oblitum*, and *L. arenosus* a *nomen protectum*. The species is redescribed, its features are illustrated, and its distribution extends from Panama to Chile, in shallow subtidal to platform depths in the Eastern Pacific.

Keywords: Synonymy; Reversal of precedence; *Lepidonotus*; Bidentate neurochaetae

Resumen

Émile Blanchard describió 3 especies de polinóidos de Chile: *Polynoe chilensis*, *P. fasciculosa* y *P. virens*. Se encontró el material tipo de *P. chilensis* en las colecciones del Museo Nacional de Historia Natural de París, pero las otras 2 especies han sido consideradas como indeterminables y carecen de material tipo. Durante una estancia de investigación en París, se hallaron los ejemplares tipo de *P. fasciculosa* y en comparación con otros materiales depositados en el Museo de Historia Natural del Condado de Los Angeles, se concluyó que la especie es un sinónimo de *Lepidonotus arenosus* Ehlers, 1901, ya que posee 12 pares de élitros, antenas laterales terminales, élitros con

fimbria, macro y microtubérculos globulares a polyhédricos, y neurosetas apenas bidentadas, con los denticulos accesorios, a menudo erosionados. No obstante, ya que *P. fasciculosa* sea un homónimo menor de *P. fasciculosa* Grube, 1840, o combinado como *Lepidonotus fasciculosus*, se le considera como *nomen oblitum*, a *L. arenosus* como *nomen protectum*. Se redescrive a la especie, se ilustran sus atributos y se extiende su distribución de Panamá a Chile, en fondos someros a profundos en la plataforma continental en el Pacífico oriental.

Palabras clave: Sinonimia; Reversión de precedencia; *Lepidonotus*; Neurosetas bidentadas

Introduction

During the revision of the species of some genera of polynoid polychaetes, the type material of 1 species described from Chile by Blanchard (1849) was found in the general collection in the Muséum National d'Histoire Naturelle, Paris. The study of these specimens allowed to conclude they belong in *Lepidonotus* and to clarify their status, this research is undertaken, which matches Christopher Kemp's idea of making expeditions in the collections of natural history museums (Salazar-Vallejo, 2018).

Charles Émile Blanchard (1819-1900) was the author of the first species of polychaetes described for the continental portion of America, since Pallas (1766) had described 2 species from Antigua, Caribbean Sea. Blanchard made his contribution to the third volume on Zoology of Claudio Gay's 30 volumes "Historia Física y Política de Chile" where 15 new species were briefly described (Blanchard, 1849).

Claudio Gay collected the specimens mostly in Chiloé (Ancud, Castro) and Calbuco; he carried up the specimens to Paris where Blanchard studied them and briefly described 15 species with Latin diagnoses and descriptions in Spanish. It is enigmatic why de Quatrefages was not asked to study the Chilean specimens, because he was in the same lab, and had already made some descriptions of new polychaete species (de Quatrefages, 1843). Blanchard started studying insects, then moved to helminths, and made several contributions regarding their nervous system and systematic affinities; in his mature life he also diversified into agriculture pests, vertebrates, geology, and how to improve teaching activities (Bouvier, 1900). Anyway, only 5 of the species names introduced by Blanchard are regarded as valid, whereas the other 10 have been regarded as indeterminable (Rozbaczylo, 1985; Rozbaczylo et al., 2017).

Blanchard described 3 polynoid species under *Polynoe*: *P. chilensis*, *P. virens*, and *P. fasciculosa*. The diagnosis indicated that *P. chilensis* had 14 pairs of large tuberculate elytra (Blanchard, 1849), leaving middorsal area uncovered (Atlas, Pl. 1, Fig. 1), *P. virens* had 35 pairs of small elytra (Blanchard, 1849), the small elytra leave

most dorsal surface uncovered (Atlas, Pl. 1, Fig. 2), and *P. fasciculosa* had 14 pairs of large tuberculate elytra (Blanchard, 1849), not covering middorsal area (Atlas, Pl. 1, Fig. 3).

Baird thought that *P. chilensis* and *P. virens* might belong in *Halosydna* Kinberg, 1856 (Baird, 1865), whereas *P. fasciculosa* might belong in *Antinoe* Kinberg, 1856 (Baird, 1865). For the former species he noted: "the figures do not correspond with the descriptions. In *Polynoe chilensis*, M. Blanchard describes the number of elytra as being 16 pairs, "covering the back entirely" (quotation marks in the original). In his figure he depicts the animal as having 29 pairs of elytra, which leave the back naked in the centre! In *P. virens* he describes the animal as having 35 pairs of elytra, while he figures it as possessing 56 or 57!"

Blanchard species were not included by de Quatrefages (1866) nor by Grube (1870). However, de Quatrefages (1866) redescended *P. chilensis* and renamed it as *P. chilensis*; he grouped this species with others having 18 pairs of elytra covering the whole dorsum. Not enough details about the insertion of lateral antennae were provided, however, such that its affinity to polynoid subfamilies or genera remain undefined.

On the other hand, Augener (1924) regarded *P. virens* as a species of *Lepidametria*. The status of the 2 other species was regarded as indeterminable. Hartman (1959) regarded *P. chilensis* and *P. fasciculosa* as indeterminable, and the latter also as a junior homonym of *P. fasciculosa* Grube, 1840 (Hartman, 1959). The latter species, however, was regarded as indeterminable by Barnich and Fiege (2009). Wesenberg-Lund (1962) regarded *P. chilensis* and *P. fasciculosa*, both described by Blanchard (1849) as indeterminable (Read & Fauchald, 2026b). Rozbaczylo (1985) listed 3 species of *Lepidonotus* Leach, 1816: *L. arenosus* Ehlers, 1901, *Lepidonotus furcillatus* Ehlers, 1901, and *Lepidonotus versicolor* Ehlers, 1901, but the latter belongs in *Thormora* Baird, 1865.

Solís-Weiss et al. (2004) included *P. chilensis* but attributed it to de Quatrefages instead of indicating it was described by Blanchard. Therefore, the correct full name for this species is *P. chilensis* Blanchard, 1849; the single letter difference between these 2 names, being based upon

the same geographic area, is not enough as to be regarded as different species (ICZN 1999, Art. 58), and they are actually the same species.

In this contribution, the type specimens of *P. fasciculosa* Blanchard (1849) are reported after being found in the general collections of the Muséum National d'Histoire Naturelle, Paris, and they have been studied for finding out about their characteristics and affinities.

Materials and methods

Specimens were observed in stereo microscopes, and parapodia and chaetae were observed in compound microscopes. Series of digital photos were stacked with HeliconFocus 8, and figures were assembled with Paint Shop Pro 2021. Specimens are deposited in the Muséum National d'Histoire Naturelle, Paris (MNHN), and in the Allan Hancock Polychaete collection of the Los Angeles County Natural History Museum, U.S.A. (LACM AHF).

Description

Order Phyllodocida Dales, 1962

Suborder Aphroditiformia Levinsen, 1883

Family Polynoidae Kinberg, 1856

Subfamily Lepidonotinae Willey, 1902

Lepidonotus Leach, 1816

Type species. Aphrodita clava Montagu, 1808, by monotypy.

Diagnosis (modified after de Assis et al., 2015). Lepidonotinae with body with 26 segments; lateral antennae terminal; antennae and cirri smooth. Facial tubercle present; buccal segment with or without nuchal fold. Twelve pairs of elytra fimbriate or with smooth margins, on segments 2, 4, 5, 7.... 21 and 23. Notopodia small or vestigial; notochaetae short, slender, spinose, sometimes capillaries. Neuropodia large, with or without acicular lobe; neurochaetae stout, long, with pectinate area with denticles, tips uni- or bidentate.

Remarks. *Lepidonotus* includes over 70 valid species (Read & Fauchald, 2026a), distributed in all oceans, mostly in shallow waters.

Lepidonotus arenosus Ehlers, 1901

Figures 1-4

Polynoe fasciculosa Blanchard, 1849, Pl. 1, Fig. 3 (*non P. fasciculosa* Grube, 1840); junior homonym.

Lepidonotus arenosus Ehlers, 1901a; Ehlers, 1901b, Pl. 2, Figs. 9-12; Seidler, 1924; Rozbaczylo, 1985; Hartwich, 1993; de Assis et al., 2015.

Lepidonotus furcillatus: Hartman, 1939 (*non* Ehlers, 1901).

Diagnosis. *Lepidonotus* with double dorsal tubercles in chaetiger 2; elytra with fimbriae, macrotubercles and microtubercles irregular, globular to polyhedral; notochaetae tapered, denticulate; neurochaetae with pectinate area, tips barely bidentate.

Description. Smaller syntype of *P. fasciculosa* (MNHN IA 2000-2134), complete, brownish, elytra overlapping laterally and dorsally, completely covering dorsum; elytra with darker macrotubercles larger along first 5 pairs of elytra, and in pairs 11 and 12 (Fig. 1A, B); median segments with smaller macrotubercles; dorsum and venter pale; elytron 5 (more oval), and 6 (paramecium like), and right parapodium of chaetiger 10 removed for observation (kept in container); body 24 mm long, 6.0 mm wide, 26 chaetigers.

Prostomium roughly hexagonal; ceratophore of median antenna about twice wider than laterals, positioned at same level as laterals (Fig. 1C); median ceratostyle subdistally swollen, tip thin, long; lateral antennae about as long as prostomium, slightly longer than palps, antennae subdistally swollen, tips thin, long. Palps smooth, tips mucronate. Eyes colorless, of similar size, anterior eyes in widest prostomial area, posterior eyes in posterior prostomial area, not covered by following segment.

Facial tubercle not seen dorsally. Tentacular segment indistinct from second chaetiger, without chaetae; left tentacular cirri lost, right dorsal and ventral ones on site, resembling antennae, as long as palps.

All parapodia biramous (Fig. 1D); dorsal cirri resembling lateral antennae, slightly wider subdistally, with tips long, tapered, dorsal cirrophore about twice longer than wide, cirrostyle with thin blunt tips, surpassing neurochaetal tips.

Second or buccal segment without nuchal lappet; 1 pair of dorsal tubercles, depressed; a single round middorsal tubercle in chaetiger 3; ventral cirri about 3 times as long as following ones, tapered, mucronate. Nephridial lobes globose, truncate, present along chaetigers 8 to 25. Notopodia reduced, low lobes; notacicular lobes digitate. Notochaetae 12-14 per bundle, most covered by foreign material, with series of rows of denticles along most of its surface, tips bare, sharp. Neuropodia with truncate prechaetal lobe, and round postchaetal lobe. Neurochaetae 20-22 per bundle, straight, barely falcate, wider than notochaetae, pectinate area swollen subdistally, with series of denticles, progressively larger distally, tips unidentate, accessory denticles eroded (Fig. 1E).

Posterior region tapered; anus dorsoterminal in last chaetiger, anal cirri lost.

Elytra all oval, longer than wide, fimbriate along posterior margins; elytra surface with macro- and microtubercles spread along surface, concentrated in a

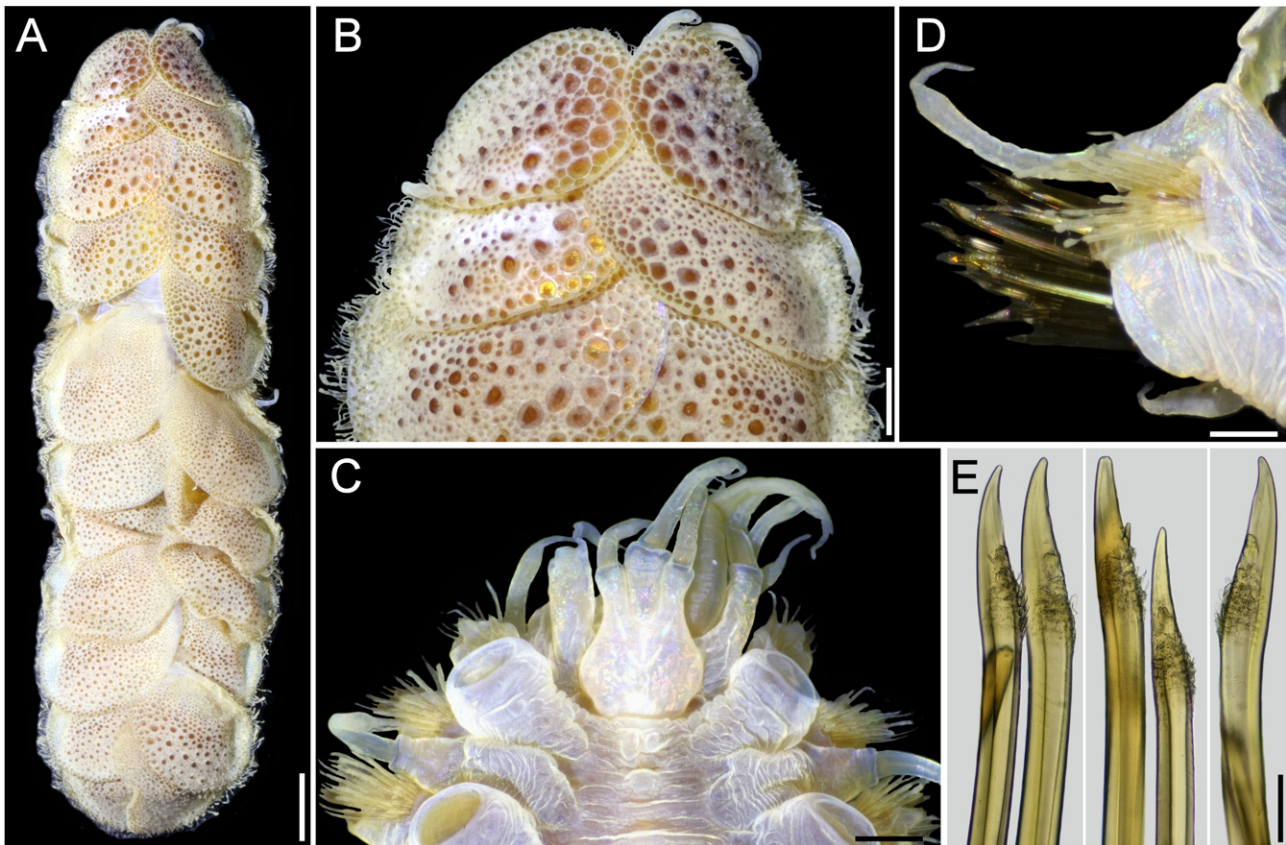


Figure 1. *Lepidonotus arenosus* Ehlers, 1901, syntype of *Polynoe fasciculosa* Blanchard, 1849 (MNHN IA 2000-2134). A, Dorsal view; B, anterior end, dorsal view; C, same, elytra removed, dorsal view; D, chaetiger 10, right parapodium, anterior view; E, same, upper, median and lower neurochaetae. Scale bars: A, 2 mm; B, 0.9 mm; C, 0.6 mm; D, 0.4 mm, E, 70 μ m.

large central area, leaving a wide band along posterior region without macrotubercles (Fig. 2A); macro- and microtubercles globose to polyhedral, often with granulose surface. Anterior area only with microtubercles, leaving margins bare (Fig. 2B). Inner median area with macro- and microtubercles, margin with tiny globular papillae (Fig. 2C, inset). Outer posterior area with fimbriae and microtubercles, without macrotubercles (Fig. 2D). Median posterior area fimbriate with macrotubercles and microtubercles, leaving a wide band without macrotubercles (Fig. 1E). Insertion area with macro- and microtubercles, some with a distinct tip (Fig. 2F).

Variation. The additional specimens (LACM AHF 429-35, 863-38) were fragments, but their morphological features correspond with the syntypes because their elytra are fimbriate, having macro- and microtubercles, and have a wide band along posterior area without macrotubercles.

The specimens are in suboptimal condition with most body-wall transparent (Fig. 3A). The anterior eyes retain some pigmentation and are in the widest prostomial

area (Fig. 3B). The notochaetae are cleaner and their denticulation and sharp long tips are better defined (Fig. 3C, D). Neurochaetae show gradation in the erosion of accessory denticles and in the size of spines along the pectinate area (Fig. 3E), the posterior region is completely covered by elytra, and the pygidium has an anal plate with 2 terminal anal cirri (Fig. 3F, G).

Elytra are oval, longer than wide (Fig. 4A). The external median area has only microtubercles and smooth margin (Fig. 4B). The insertion area has macro- and microtubercles globular, some with a barely defined tip (Fig. 4C). The anterior inner area has macro- and microtubercles polyhedral or globose (Fig. 4D). The external posterior area is fimbriate with microtubercles teardrop shaped (Fig. 4E). The inner posterior area has tear-drop macro- and microtubercles, without fimbriae (Fig. 4F).

Taxonomic summary

Type material. Chile. Two syntypes of *P. fasciculosa* Blanchard, 1849 (MNHN IA 2000-2134), collected

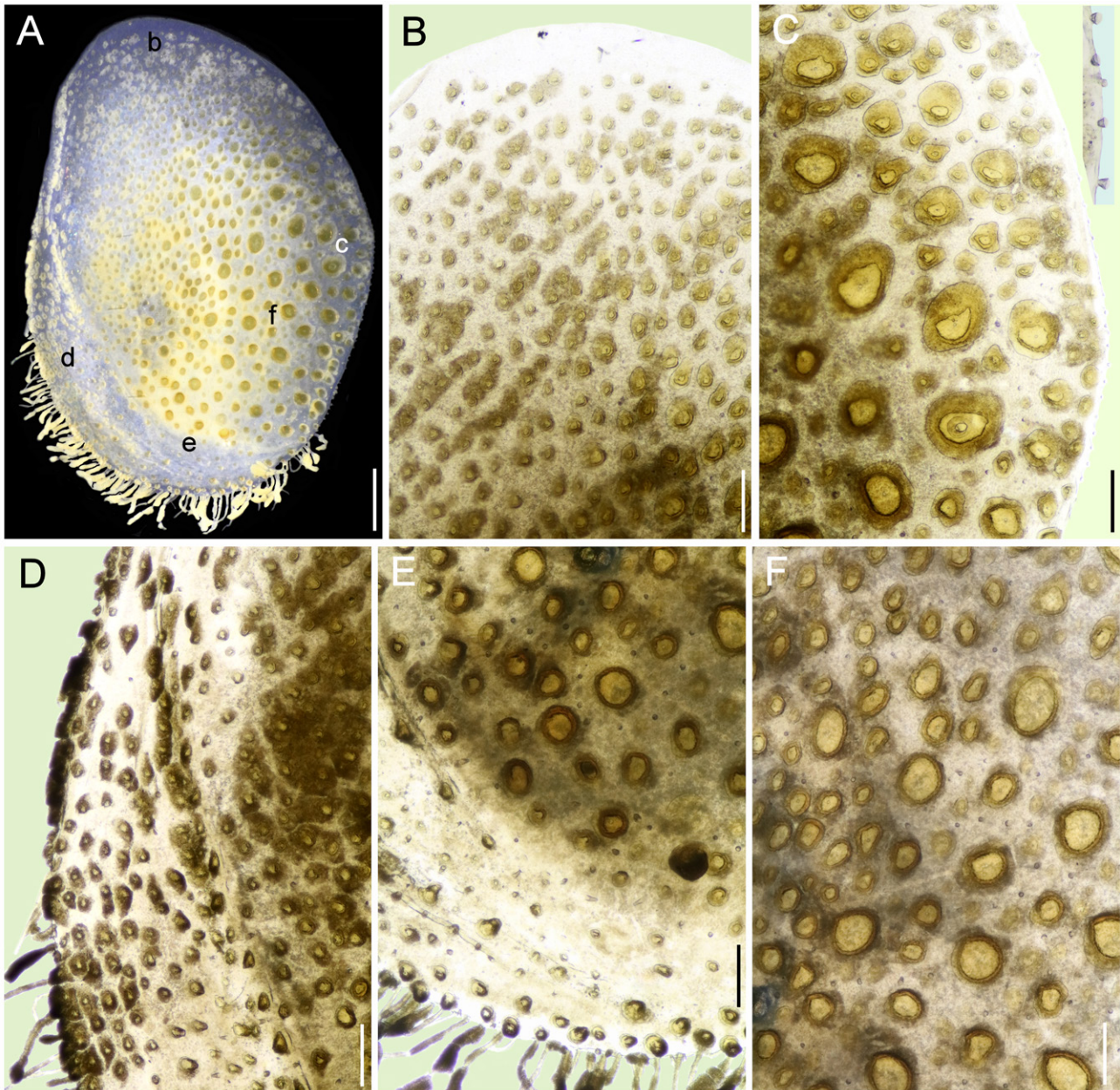


Figure 2. *Lepidonotus arenosus* Ehlers, 1901, syntype of *Polynoe fasciculosa* Blanchard, 1849 (MNHN IA 2000-2134). A, Left fifth elytron, seen from above (letters indicate enlarged areas); B, anterior covered area with microtubercles; C, inner median area with macro- and microtubercles, margin with tiny cup-shaped papillae (inset: close-up of papillae); D, external posterior area with long fimbriae, and marginal microtubercles; E, posterior median area with fimbriae, marginal microtubercles and macro- and microtubercles; F, insertion area with macro- and microtubercles. Scale bars: A, 0.6 mm; B-F, 0.2 mm.

in Calbuco, Jan. 1836 (after Muñoz-Pizarro 1944; the syntypes are in different condition, the smaller is better preserved than the larger one, and the former was used for making photographs; larger syntype 29 mm long, 6.5 mm wide, 26 chaetigers).

Additional material. Colombia. One specimen (LACM AHF 429-35), anterior fragment, RV Velero III, Sta. 429-35 (06°48'40" N, 77°40'50" W), Bahía Octavia, 63-54 m, coarse sand, gravel; 1 Jan. 1935 (mature female, oocytes 100 µm in diameter; left elytron 5, and left parapodium of

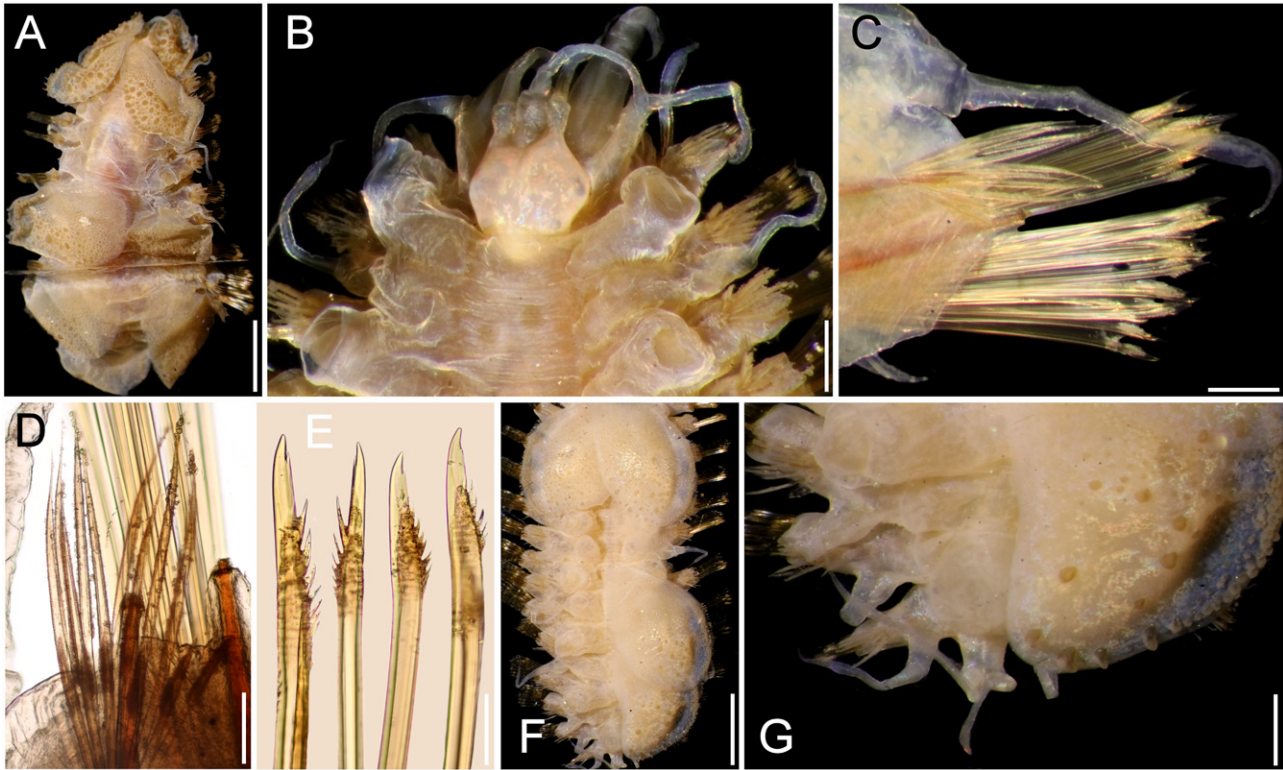


Figure 3. *Lepidonotus arenosus* Ehlers, 1901, non-type specimens. A, Anterior fragment (LACM 429 35), depressed by glass slide, dorsal view; B, anterior end, after removal of elytra; C, chaetiger 10, left parapodium, anterior view; D, same, notochaetae; E, same, tips of upper, median and lower neurochaetae; F, posterior fragment (LACM 863 38), dorsal view; G, same, close-up of posterior end. Scale bars: A, 2.2 mm; B, 0.6 mm; C, 0.3 mm; D, 0.5 mm; E, 80 μ m; F, 1.8 mm; G, 0.7 mm.

chaetiger 10 removed for observation (kept in container); fragment 11 mm long, 6 mm wide, 13 chaetigers; nephridial lobes from chaetiger 8). Panama. One specimen (LACM AHF 863-38), posterior fragment, RV Velero III, Sta. 863-38 (07°45'35" N, 81°35'35" W), Bahía Honda, 54-90 m, rock, sand, mud; 3 Jan. 1939 (posterior fragment, some elytra already detached; high basal tubercles in dorsal cirri; posterior elytron of unknown chaetiger, and left elytron 12; fragment 14 mm long, 6 mm wide, 14 chaetigers; nephridial lobes to last chaetiger; anus in chaetiger 25, anal plate longer than wide, with long anal cirri). Ecuador. One specimen (LACM 171-34), broken into anterior and posterior fragments, RV Velero III, Sta. 171-34 (00°46'10" S, 89°30'10" W), off Stephens Bay, Chatham Island, Galápagos, 63-72 m, fine sand, corallines, 21 Jan. 1934 (anterior fragment 14 mm long, 6 mm wide, 13 chaetigers; posterior fragment 9 mm long, 5 mm wide, 13 chaetigers; nephridial lobes along chaetigers 8-25).

Distribution. Eastern Pacific, Panamá to Chile, in mixed bottoms, intertidal to shallow subtidal in temperate areas, in platform depths in tropical areas.

Remarks

The printed label in the syntype vial indicates "*Polynoe fasciculosa* Blainv., M. Gay, Chili, A1(R.) - 1868- No. 30a". The author of the species is wrong and it can be explained because the handwritten label might have had the last name abbreviated, and it was confused with Blainville. This later author made several publications dealing with polychaetes but only in 1816-1828, 20 years before the publication of the chapter by Blanchard (1849). The collector and country are correct, and the year in the label must correspond to the time when the label was printed.

Polynoe fasciculosa Blanchard, 1849 is a primary junior homonym of *P. fasciculosa* Grube, 1840, as indicated elsewhere (Read & Fauchald, 2026b), and must be replaced (ICZN, 1999, Art. 60). The syntypes belong in *Lepidonotus* Leach, 1816 by having lepidonotin prostomium, 12 pairs of elytra, elytra overlapping laterally and dorsally, parapodia without branchiae, tapered denticulate notochaetae, and neurochaetae with pectinate area. Consequently, it could be newly combined as *L. fasciculosus* (Blanchard, 1849).

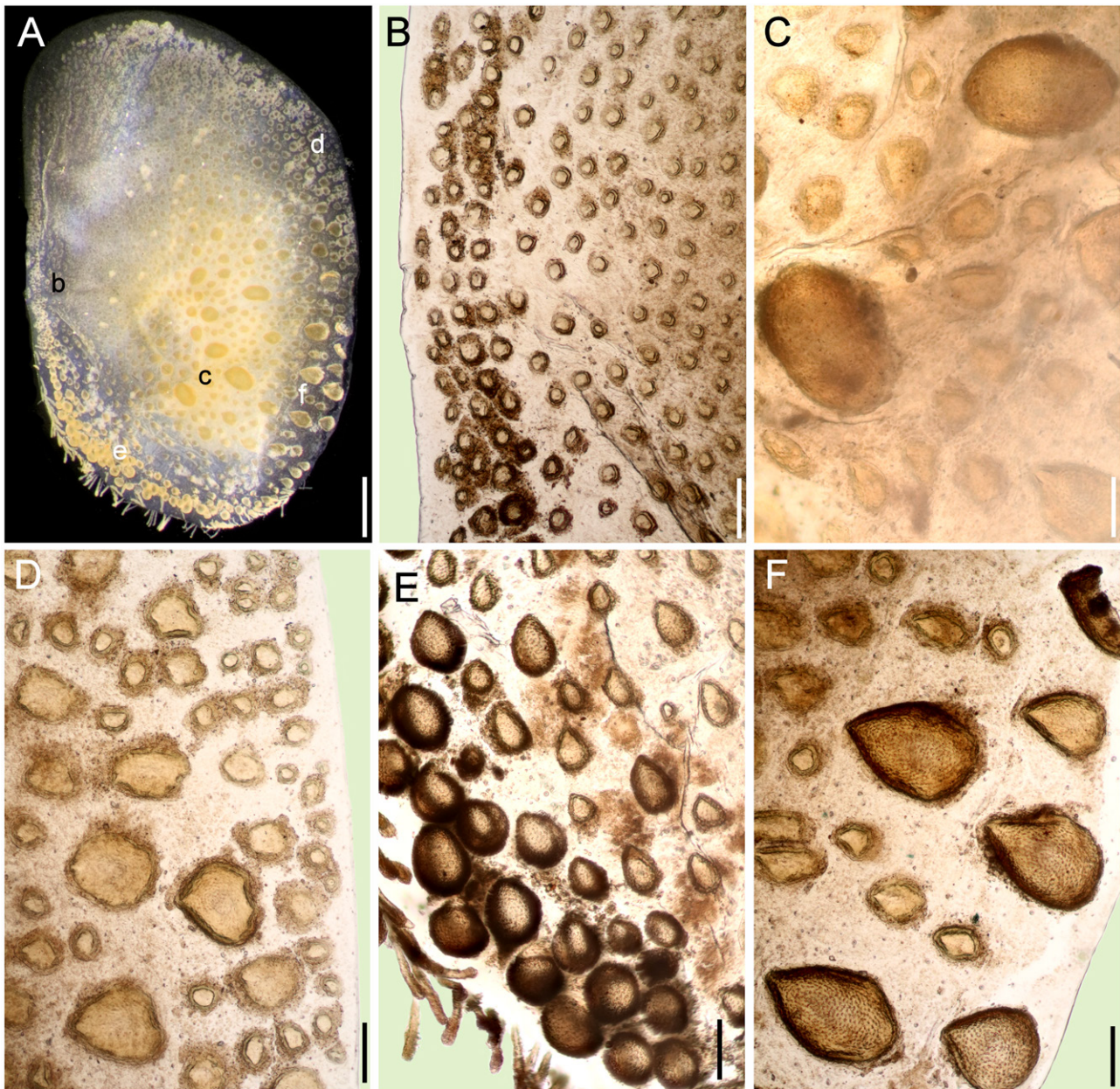


Figure 4. *Lepidonotus arenosus* Ehlers, 1901, non-type specimen (LACM 429 35). A, Left elytron 5, seen from above (letters indicate enlarged areas); B, anterior external area with globular microtubercles; C, insertion area with largest oval or tear-drop shaped macro- and microtubercles; D, posterior interior area with polyhedral macro- and microtubercles; E, external fimbriate margin and adjacent area with teardrop shaped macro- and microtubercles; F, posterior smooth margin and adjacent area with teardrop shaped macro- and microtubercles. Scale bars: A, 0.5 mm; B-F, 70 μ m.

Barnich and Fiege (2009) regarded the senior homonym, *P. fasciculosa* Grube, 1840 as indeterminable after an insufficient description and the lack of type material. Consequently, because the identity of the senior homonym cannot be confirmed, these 2 names cannot be regarded as congeneric and the secondary homonym

would not need to be replaced (ICZN, 1999, Art. 59.2). Thus, the junior homonym could be retained and newly combined as *L. fasciculosus* (Blanchard, 1849).

It must be noted, however, that *L. fasciculosus* resembles *L. arenosus* Ehlers, 1901a, also described from Calbuco, Chile, and redescribed by Ehlers (1901b, Figs. 9-12).

Both species have similar prostomia, dorsal tubercles, and elytra fimbriate with large, polyhedric macro- and microtubercles. The only potential difference would be the type of neurochaetae. Ehlers (1901b) described and illustrated neurochaetae as having progressively larger denticles in the pectinate area, and tips bidentate. The syntypes of *L. fasciculosus* also have a similar pectinate area, but tips are unidentate. However, the neurochaetae are badly eroded and the difference can be regarded as an effect of erosion, and not a true difference between the species. Therefore, these 2 species are regarded as synonyms and *L. fasciculosus* would have priority.

Nevertheless, because the status of *P. fasciculosa* was regarded as indeterminable as early as 1865 (Baird, 1865), it has not been used as a valid name, and using *L. fasciculosus* over *L. arenosus* would contradict the current general use because *L. arenosus* is an accepted name. Further, a reversal of precedence is not appropriate because ‘the senior homonym has not been used as a valid name after 1899’ (ICZN, 1999, Art. 23.9.1.1), and the second requirement of the number of authors and publications might be difficult to fulfill because there are few taxonomists working in the area, the species is apparently not too abundant or ecologically relevant, and there might be less than the 25 publications by 10 authors during the last 50 years (ICZN, 1999, Art. 23.9.1.2). However, in any faunistic or ecological publication about Chilean polynoids, *L. arenosus* is the name used, not its senior synonym (Rozbaczylo et al., 2017). Therefore, *P. fasciculosa*, or its newly combined *L. fasciculosus* must be regarded as the *nomen oblitum*, and *L. arenosus* the *nomen protectum* (ICZN, 1999, Art. 23.9.2).

On the other hand, Augener (1913) regarded *L. arenosus* and *L. furcillatus* Ehlers, 1901 as synonyms, and Hartman (1939) followed him even though she found differences in elytral macro-tubercles. These 2 species have fimbriate elytra with macro- and microtubercles, and neurochaetae with tips bidentate, and the additional denticle is very small. However, Seidler (1924) regarded them as different species because in *L. arenosus* there is no nuchal lappet, elytra do not have innervation, and macro-tubercles are globular to polyhedric, whereas in *L. furcillatus* the nuchal lappet has 2 blunt conical protuberances, elytra have innervation, and macro-tubercles are conical or bifid. Further, Augener’s record of *L. furcillatus* for New Zealand deserves confirmation. Likewise, Pruvot’s (1930; repeated by Fauvel, 1947) record of *L. arenosus* for New Caledonia also deserves confirmation; there are some differences because the 2 specimens Pruvot had studied in comparison to typical *L. arenosus*. For example, New Caledonian specimens have anterior eyes displaced posteriorly (in the widest area in *L. arenosus*), macro-tubercles are in a

smaller elytral surface and their shape is globular (against being extended over a larger area, and with irregular to polyhedric shape in *L. arenosus*), and micro-tubercles have distinct spines (missing in *L. arenosus*). If these features are confirmed, then it might belong to a different, probably undescribed species.

Discussion

The type material of *P. fasciculosa* Blanchard, 1849 was collected in 1836; they were deposited in the Muséum National d’Histoire Naturelle, Paris, in the early 1840s and described less than 20 years after collection. Regretfully, Blanchard polynoid species were regarded as indeterminable by Baird (1865), and they were not incorporated in full by de Quatrefages (1865) because he only included *P. chilensis*, as indicated above. The finding of the type material did not lead to the reinstatement of an old species name because of the formalities and regulations of the International Code of Zoological Nomenclature (1999); its status was clarified as an earlier synonym of *L. arenosus* Ehlers, 1901, but the earlier name must be regarded as a *nomen oblitum*, and the latter a *nomen protectum*.

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